



# scale-X™ bioreactor for viral vector production

Proof of concept for scalable and homogeneous  
Vero cell growth

## Application note

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## Abstract

The scale-X™ bioreactor is a novel fixed-bed system that enables cultivation of surface-dependent and suspension cell lines. A large surface area in small-footprint units enable high cell concentrations. This study compared adherent Vero cell culture in scale-X hydro bioreactors with standard microcarrier cultivations. Both systems enabled similar surface-specific cell concentrations. Subsequent unwinding of the spiral-wound fixed-bed revealed a uniform cell distribution in axial and radial directions. This indicates a homogeneous fluid flow and cell attachment inside the bioreactor. The observed homogenous cell growth is an important scaling factor and allows maximum use of available surface area in a reproducible manner.

## Introduction to the scale-X bioreactor

Most viral vaccines manufacturing processes are based on surface-dependent cell platforms. Typical cultivation systems such as roller bottles, multi-tray cell culture dishes and microcarrier bioreactors have constraints for large-scale production. To overcome potential scale-up limitations and to ease manufacturing, fixed-bed bioreactors are a valuable alternative. The innovative scale-X bioreactor is a scalable fixed-bed system from research- (2.4 m<sup>2</sup> of growth surface) to pilot- (10 and 30 m<sup>2</sup> of growth surface) and manufacturing-scale (200 and 600 m<sup>2</sup> of growth surface; Fig. 1). In comparison to randomly packed polyethylene terephthalate (PET) fabric strips, the proprietary scale-X technology uses a structured, spiral wound PET fixed-bed to reduce local over compaction and dead zones, while making optimal usage of available surface area with high batch-to-batch reproducibility (Fig. 2).

## Materials, Methods & Equipment

### Fixed-bed process: scale-X hydro bioreactor

#### Cell line and process conditions

|                        |   |
|------------------------|---|
| Cell line              | Vero ATCC® CCL-81.5TM                   |
| Seeding concentration  | 1×10 <sup>4</sup> cells/cm <sup>2</sup> |
| Reactor volume         | 700 mL                                  |
| Fixed-bed volume       | 170 mL                                  |
| Fixed-bed surface area | 2.4 m <sup>2</sup>                      |
| Working volume         | 2700 mL                                 |
| Stirring speed         | 740 rpm                                 |

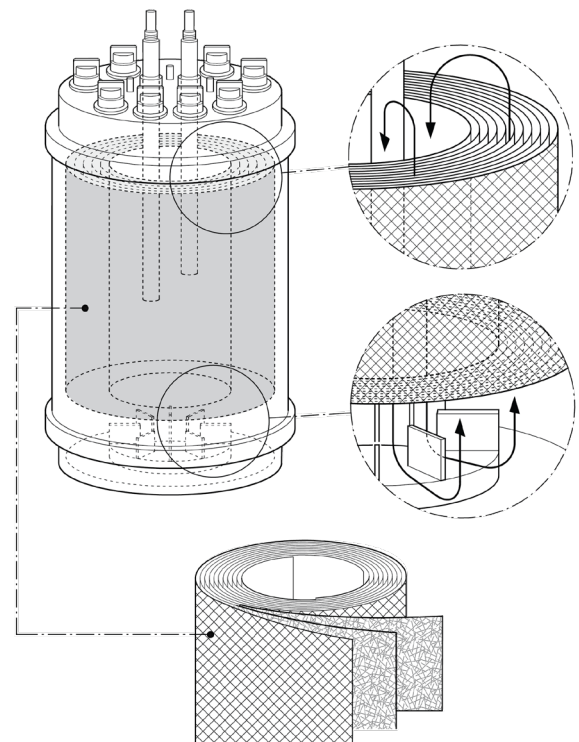
#### Method for evaluation of cell concentration and distribution

- ▶ Twelve incorporated sample strips (each 12.15 cm<sup>2</sup>) were extracted from the fixed-bed and well mixed for 3 min in 1 mL cell lysis buffer. Released nuclei were stained using crystal violet and manually counted.
- ▶ When the targeted cell concentration is reached, the fixed-bed was removed, carefully rolled out and defined areas of 1 cm<sup>2</sup> were cut (Fig. 3). Cells from the strip were lysed with reagent A100 (ChemoMetec) and the nuclei were counted with a hemocytometer.

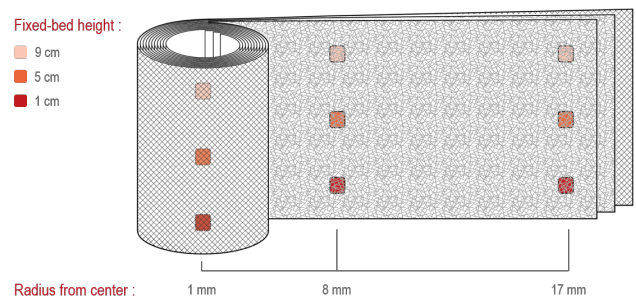
scale-X™



**Figure 1: scale-X bioreactor range from hydro (2.4 m<sup>2</sup> for R&D) to carbo (10 and 30 m<sup>2</sup> for pre-clinical) and nitro (200 and 600 m<sup>2</sup> for commercial).**



**Figure 2: Proprietary fixed-bed structure and media flow path in the scale-X bioreactor series.**



**Figure 3: Sampling points for cell concentration measurements in the scale-X fixed-bed.**

## Reference process: Cytodex® 1 microcarriers in stirred-tank bioreactor (STR)

### Cell line and process conditions

|                                   |                                       |
|-----------------------------------|---------------------------------------|
| <b>Cell line</b>                  | Vero ATCC® CCL-81.5                   |
| <b>Seeding concentration</b>      | $2 \times 10^4$ cells/cm <sup>2</sup> |
| <b>Reactor volume</b>             | 850 mL                                |
| <b>Microcarrier concentration</b> | 2 g/L Cytodex® 1                      |
| <b>Microcarrier surface area</b>  | 0.75 m <sup>2</sup>                   |
| <b>Working volume</b>             | 850 mL                                |
| <b>Stirring speed</b>             | 90 rpm                                |

### Method for cell concentration measurement

- ▶ A sample volume of 5 mL was collected from the bioreactor. After three washing steps, cells were disrupted with a 0.1 M citric acid, 0.1% crystal violet and 0.1% Triton X-100 solution.
- ▶ After an incubation time of 1 h at 37°C, stained nuclei were counted with a hemocytometer. The method is described in detail by Trabelsi et al. (Khaled Trabelsi, 2004)

### Process parameters (for both cultivations)

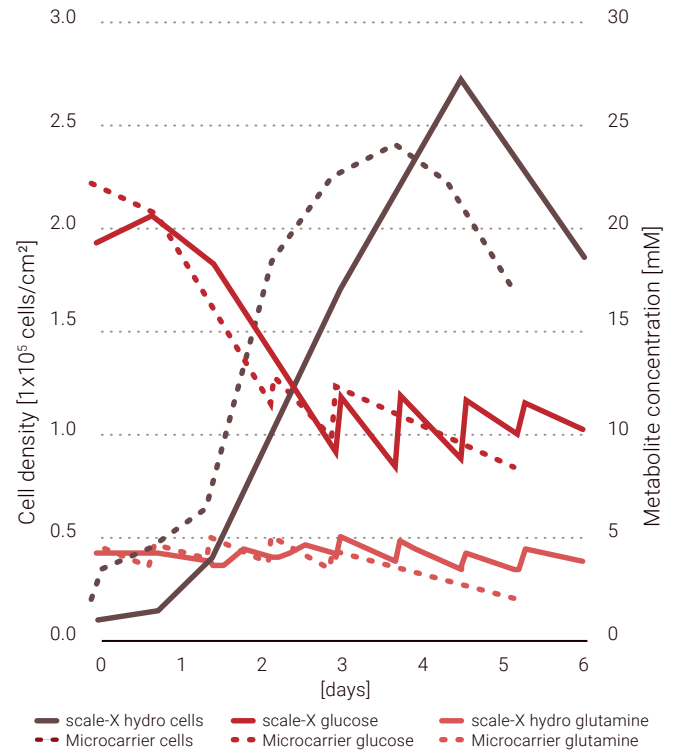
- ▶ Serum-free medium (VP-SFM, Gibco™) supplemented with 4 mM glutamine
- ▶ Feeding of glucose and glutamine stock solutions to prevent substrate limitation below 8 mM and 2 mM, respectively
- ▶ pH control at 7.2 with CO<sub>2</sub> and 0.5 NaHCO<sub>3</sub> (microcarrier) or 0.5 NaOH (scale-X)
- ▶ Temperature set point at 37°C
- ▶ Dissolved Oxygen set point at 50% (headspace aeration)

## Vero cell growth

Comparison of maximum Vero cell concentrations in different bioreactor systems (Kiesslich, 2020) (Fig. 4):

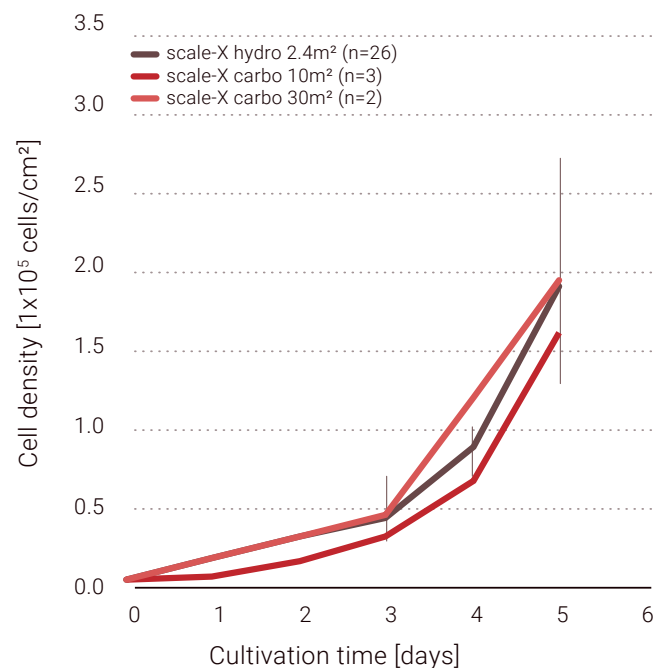
**scale-X hydro bioreactor (2.4 m<sup>2</sup>):**  $2.7 \times 10^5$  cells/cm<sup>2</sup>  
**STR Cytodex® 1 (0.75 m<sup>2</sup>):**  $2.4 \times 10^5$  cells/cm<sup>2</sup>

Different cell seeding concentrations resulted in shifted but comparable growth profiles. Surface-specific cell concentrations (Fig. 4) were slightly higher in the scale-X bioreactor ( $2.7 \times 10^5$  cells/m<sup>2</sup>) compared to the microcarrier cultivation ( $2.4 \times 10^5$  cells/m<sup>2</sup>). Both systems allowed a similar total medium usage with  $2.1$ – $2.2 \times 10^9$  cells/L<sub>medium</sub>. When considering the reactor volume of both systems at equivalent throughput, the fixed-bed bioreactor represents a strongly reduced equipment footprint (5-fold reduction).



**Figure 4: Vero cell growth in a scale-X hydro bioreactor compared to a standard microcarrier cultivation, kindly provided by Kiesslich et al. (2020).**

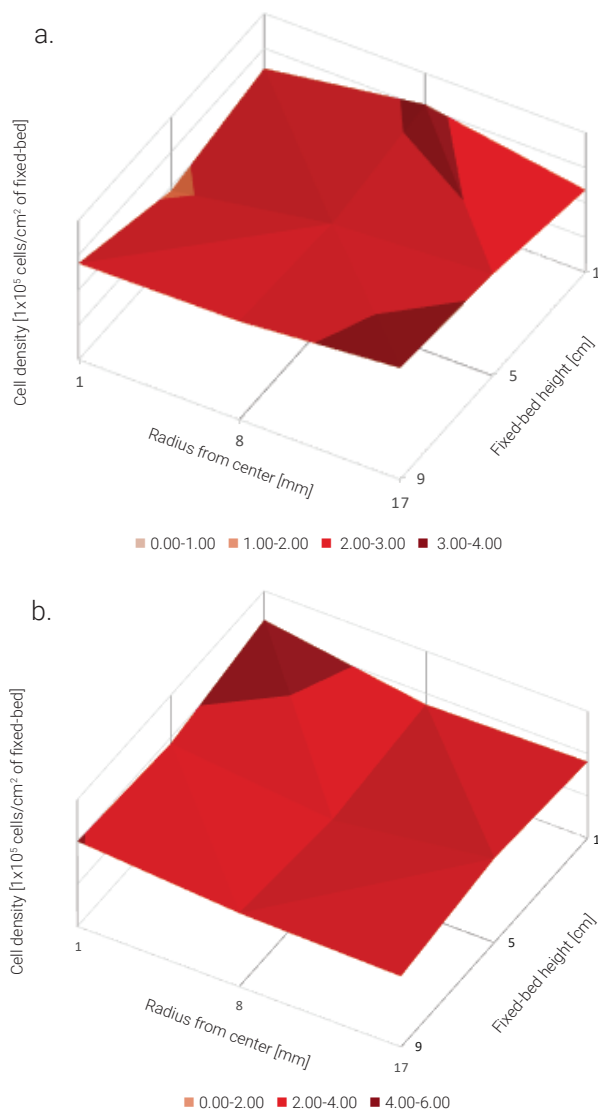
An extended Vero cell growth study was done at Univercells (Fig. 5). Multiple experiments were performed in the scale-X hydro bioreactor (n=26) resulting in reproducible cell concentrations of an average of  $2.5 \times 10^5$  cells/cm<sup>2</sup>. The scale-up to the larger scale-X carbo bioreactor (10 m<sup>2</sup> and 30 m<sup>2</sup>) resulted in similar surface-specific cell concentrations



**Figure 5: Repeated growth studies with Vero cells in different scale-X bioreactor systems, performed at Univercells.**

## Vero cell distribution in a structured, spiral wound fixed-bed

Vero cell distribution was investigated at different stages of the cultivation (at  $2.7 \times 10^5$  cells/cm<sup>2</sup> and  $5.3 \times 10^5$  cells/cm<sup>2</sup>, Fig. 6). Therefore, nine samples were extracted from the unwound fixed bed and cell concentrations were determined. Vero cells in the scale-X hydro bioreactor were homogeneously distributed among the fixed-bed, both in axial and radial direction. Additional computational fluid dynamic studies were performed and indicated the absence of dead zones and inhomogeneities in the structured fixed-bed (data not shown). Uniform cell growth enabled the full usage of available surface area



**Figure 6: Vero cell growth distribution in the structured fixed-bed of scale-X bioreactors.** Cell distribution among the radial (radius from center) and axial direction (fixed-bed height) is homogeneous. (a.) Kindly provided by Kiesslich et al. (2020), (b.) Performed at Univercells.

## Conclusion and perspectives

Adherent Vero cells reached equal concentrations in both processes. When transferred to larger fixed-bed scale-X bioreactors, similar cell growth profiles and maximum surface specific cell concentrations were achieved. Finally, studies on the Vero cell growth distribution revealed a homogenous cell attachment and growth within the fixed-bed. The compact design of the fixed-bed allows a reduction in equipment footprint by factor 5 compared to the microcarrier process. Overall, results show that the structured scale-X fixed-bed bioreactor system is a suitable alternative to traditional "scale-out" technologies such as multi-tray dishes and roller bottles, but also to technically challenging cultivation systems like microcarrier processes. The current scale-X bioreactor can be transferred to manufacturing scale (600 m<sup>2</sup>) and integrated into a continuous NevoLine™ manufacturing platform. The NevoLine system couples intensified upstream processing to continuous downstream processing to allow low-cost, large-scale capacity with reduced capital and operating costs.

## Bibliography

- Khaled Trabelsi, S. R. (2004). Comparison of various culture modes for the production of rabies virus by Vero cells grown on microcarriers in a 2-l bioreactor. *Enzyme and Microbial Technology*.
- Kiesslich, S. V.-C.-F. (2020). Serum-free production of rVSV-ZEBOV in Vero cells: Microcarrier bioreactor versus scale-X™ hydro fixed-bed. *Journal of Biotechnology*.